

Interpretation of mitochondrial diversity in terms of taxonomy: a case study of *Hyponephele lycaon* species complex in Israel (Lepidoptera, Nymphalidae, Satyrinae)

Vladimir A. Lukhtanov^{1,2}, Asya V. Novikova³

1 Department of Karyosystematics, Zoological Institute of the Russian Academy of Sciences, Universitetskaya nab. 1, 199034 St. Petersburg, Russia **2** Department of Entomology, Faculty of Biology, St Petersburg State University, Universitetskaya nab. 7/9, 199034 St. Petersburg, Russia **3** Department of Ecology, Evolution and Behavior, the Hebrew University of Jerusalem, Givat Ram, Berman bldg, 91904 Jerusalem, Israel

Corresponding author: Vladimir Lukhtanov (lukhtanov@mail.ru)

Academic editor: V. Kuznetsova | Received 26 September 2015 | Accepted 8 November 2015 | Published 19 November 2015

<http://zoobank.org/A57815EA-F70D-45A9-8762-0CD1ECB2AB99>

Citation: Lukhtanov VA, Novikova AV (2015) Interpretation of mitochondrial diversity in terms of taxonomy: a case study of *Hyponephele lycaon* species complex in Israel (Lepidoptera, Nymphalidae, Satyrinae). In: Lukhtanov VA, Kuznetsova VG, Grozeva S, Golub NV (Eds) Genetic and cytogenetic structure of biological diversity in insects. ZooKeys 538: 21–34. doi: 10.3897/zookeys.538.6689

Abstract

It is difficult to interpret mitochondrial diversity in terms of taxonomy even in cases in which a concordance exists between mitochondrial, ecological and morphological markers. Here we demonstrate this difficulty through a study of Israeli *Hyponephele* butterflies. We show that samples commonly identified as *Hyponephele lycaon* are represented on Mount Hermon in Israel by two sympatric groups of individuals distinct both in mitochondrial DNA-barcodes (uncorrected *p*-distance = 3.5%) and hindwing underside pattern. These two groups were collected in different biotopes. They also tended to be different in length of brachia in male genitalia, although the latter character is variable. We reject the hypothesis that the discovered *COI* haplogroups are selectively neutral intraspecific characters. We hypothesize that they represent: either (1) two different biological species, or (2) a consequence of a strong positive selection acting at intraspecific level and resulting in two intraspecific clusters adapted to low and to high elevations. If we accept the first hypothesis, then provisionally these two haplogroups can be attributed to transpalearctic *H. lycaon* sensu stricto and to *H. lycaonoides*, previously known from Iran and East Turkey.

Keywords

adaptation to high/low elevation, biodiversity, *COI*, cryptic species, DNA barcoding, disruptive selection, habitat-related selection, molecular markers

Introduction

Hyponephele Muschamp, 1915 is a large and taxonomically diverse genus of satyrine butterflies. The genus contains 39 species (Eckweiler and Bozano 2011) distributed throughout the Palearctic region, with the highest species diversity found in Central Asia, Iran and Turkey. This group was taxonomically revised by Samodurov with co-authors (Samodurov et al. 1995, 1996, 1997, 1999a, b, 2000, 2001) and by Eckweiler and Bozano (2011).

Within the genus, *Hyponephele lycaon* (Rottenburg, [1775]) is the best known and the most common species broadly distributed in the temperate zone of the Palearctic from Portugal in the west to Far East Russia in the east (Samodurov et al. 2001, Eckweiler and Bozano 2011). In south Palearctic it is replaced by closely related allopatric taxa *H. maroccana* Blachier, 1908 (North Africa), *H. galtscha* (Grum-Grshimailo, 1893) (Tajikistan) and *H. sifanica* (Grum-Grshimailo, 1891) (China) (Eckweiler and Bozano 2011). One more species, *H. lycaonoides* D. Weiss, 1978 was described from Zagros Mountains in Iran. *Hyponephele lycaonoides* was shown to be sympatric with *H. lycaon* in Iran (Weiss 1978, Eckweiler and Bozano 2011, Tshikolovets et al. 2014). *Hyponephele lycaonoides* was also reported for Turkey (Koçak 1989, Eckweiler and Bozano 2011), but the reports for Turkey were questioned in the comprehensive analysis of Turkish butterfly fauna made by Hesselbarth et al. (1995). Male genitalia structures are commonly used for distinguishing between *H. lycaon* and *H. lycaonoides*, and specimens with long brachia are attributed to *H. lycaon*, whereas specimens with short brachia are attributed to *H. lycaonoides* (Weiss, 1978). However, male genitalia are variable in both *H. lycaon* and *H. lycaonoides*, and intermediate forms are reported to be common (Eckweiler and Bozano 2011). Moreover, Hesselbarth et al. (1995) considered these traits (the long and short brachia) as intraspecific variations, rather than species-specific characters. Unfortunately, until now nobody used molecular markers to test the non-conspecificity of *H. lycaon* and *H. lycaonoides*.

In our study we analysed mitochondrial DNA barcodes and morphological and ecological markers to show that butterflies commonly identified as *Hyponephele lycaon* are represented in Israel by two sympatric groups of individuals. We further discuss different possible evolutionary and taxonomic interpretations of the pattern discovered.

Materials and methods

In the course of our DNA barcode survey of Israeli butterflies (2012–2015) we found butterflies similar to *H. lycaon* on Mount Hermon in northern Israel. They were collected in a small area situated between 33°17'12"N, 35°45'49"E, at 1440 m and 33°18'38"N, 35°47'07"E, at 2050 m. The distance between these extreme points of the collecting was 3460 m (measured using Google Earth map). Some of the butterflies were collected in the forest zone at 1450–1600 m above sea level, other were collected

Table 1. List of *Hyponephele* samples sequenced in the present study.

Haplogroup or taxon	Country	Ecological zone	Pattern of the wing underside	BOLD Process ID	Field ID	GenBank accession #
I	Israel	forest	contrasting	BPAL2756-15	CCDB-17969_A01	KT864697
I	Israel	forest	contrasting	BPAL2757-15	CCDB-17969_A02	KT864698
I	Israel	forest	contrasting	BPAL2758-15	CCDB-17969_A03	KT864699
I	Israel	forest	contrasting	BPAL2760-15	CCDB-17969_A05	KT864700
I	Israel	forest	contrasting	BPAL2761-15	CCDB-17969_A06	KT864701
I	Israel	forest	contrasting	BPAL2765-15	CCDB-17969_A10	KT864702
II	Israel	forest	pale	BPAL2695-14	CCDB-17968_C11	KT864691
II	Israel	subalpine	pale	BPAL2705-14	CCDB-17968_D09	KT864692
II	Israel	subalpine	pale	BPAL2706-14	CCDB-17968_D10	KT864693
II	Israel	subalpine	pale	BPAL2733-14	CCDB-17968_G01	KT864690
II	Israel	subalpine	pale	BPAL2762-15	CCDB-17969_A07	KT864694
II	Israel	subalpine	pale	BPAL2763-15	CCDB-17969_A08	KT864695
II	Israel	subalpine	pale	BPAL2764-15	CCDB-17969_A09	KT864696
<i>H. lupinus</i>	Israel	n/a	n/a	BPAL2719-14	CCDB-17968_E11	KT864688
<i>H. lupinus</i>	Israel	n/a	n/a	BPAL2683-14	CCDB-17968_B11	KT864689
<i>H. maroccana</i>	Morocco	n/a	n/a	BPAL1378-12	CCDB-03030_D12	KT864703
<i>H. maroccana</i>	Morocco	n/a	n/a	BPAL1377-12	CCDB-03030_D11	KT864704
<i>H. maroccana</i>	Morocco	n/a	n/a	BPAL1376-12	CCDB-03030_D10	KT864705

in the subalpine zone with predominance of xerophytous vegetation at 1800–2050 m above sea level (Table 1).

DNA barcodes, 658 bp fragments within mitochondrial gene, *cytochrome oxidase subunit I (COI)*, were sequenced at the Canadian Centre for DNA Barcoding (CCDB, Biodiversity Institute of Ontario, University of Guelph) using standard high-throughput protocol described in deWaard et al. (2008). DNA was extracted from a single leg removed from each voucher specimen employing a standard DNA barcode glass fibre protocol (Ivanova et al. 2006). All polymerase chain reactions (PCR) and DNA sequencing were carried out following standard DNA barcoding procedures for Lepidoptera as described previously (Hajibabaei et al. 2005). Photographs of specimens used in the analysis are available in the Barcode of Life Data System (BOLD) at <http://www.barcodinglife.org/>. All voucher specimens are deposited at the Zoological Institute of the Russian Academy of Sciences and could be identified by the corresponding unique BOLD Process IDs, that were automatically generated by BOLD, and by GenBank accession numbers (Table 1).

The procedure of phylogenetic inference was described previously (Vershinina and Lukhtanov 2010, Talavera et al. 2013, Lukhtanov et al. 2014, 2015a, b). Briefly, the sequences were aligned using BioEdit version 7.1.7 software (Hall 1999) and edited manually. Phylogenetic relationships were inferred using Bayesian Inference and

the program MrBayes 3.2.2 (Ronquist et al. 2012). A GTR substitution model with gamma distributed rate variation across sites and with proportion of invariable sites was specified before running the program as suggested by jModelTest (Posada 2008). Two runs of 10,000,000 generations with four chains (one cold and three heated) were performed. Chains were sampled every 10,000 generations, and burn-in was determined based on inspection of log likelihood over time plots using TRACER, version 1.4 (available at <http://beast.bio.ed.ac.uk/Tracer>). For comparison we used additional *COI* barcodes of *Hyponephele* downloaded from GenBank (Lukhtanov et al. 2009, Dinca et al. 2011).

Butterfly photographs were taken with Nikon D810 digital camera equipped with a Nikon AF-S Micro Nikkor 105 mm lens. Genitalia photographs were taken with Leica M205C binocular microscope equipped with Leica DFC495 digital camera, and processed using the Leica Application Suite, version 4.5.0 software.

Results

During a 2012–2015 survey of Israeli fauna, *H. lycaon*-similar butterflies were found only on Mount Hermon in northern Israel. We never observed *H. lycaon*-similar butterflies in other parts of Israel, although the distantly related *H. lupinus* (Costa, 1836) was found not only in the northern, but also in central Israel. Thus, our observations support the finding that the geographic range of *H. lycaon* species complex is restricted in Israel to the northernmost part of the country (Benyamin 2002).

Molecular analysis of *H. lycaon*-similar samples (Table 1, Fig. 1) revealed two distinct mitochondrial haplogroups (I and II) that were strongly differentiated with respect to the *COI* gene. These two haplogroups differed from one another by 23 fixed nucleotide substitutions in the studied 658 bp fragment of the mitochondrial *COI* gene. When looking at the level of primary polypeptide structure, these differences translate to two fixed amino acid substitutions in the studied fragment. The minimal uncorrected *COI* *p*-distance between these two haplogroups was found to be as high as 3.5 %. *Hyponephele lupinus* from Israel was found to be closely related to *H. interposita* and distant from all the taxa of the *H. lycaon* complex.

With a single exception (female sample BPAL2695-14|CCDB-17968_C11, Fig. 1, Table 1), the representatives of these two *COI* haplogroups were collected in different biotopes (Fig. 2). The butterflies of haplogroup I were found on grassy slopes in the forest zone (1450–1600 m above sea level) (Fig. 2a). The butterflies of haplogroup II were found in steppe lands of the subalpine zone (1800–2050 m alt.), where xerophytic thorny cushion vegetation formed by *Onobrychis cornuta* and *Astragalus* species (Fabaceae) was predominant (Fig. 2b).

Standard χ^2 -test was used to distinguish between random *vs.* non-random distribution haplogroups I and II in the low (forest) and high (subalpine) zones. Empirical and expected frequencies of *COI* haplogroups I and II in low and high altitude belts were compared (Table 2). The calculated χ^2 was larger than the tabular value (9.558 *vs.* 6.635,

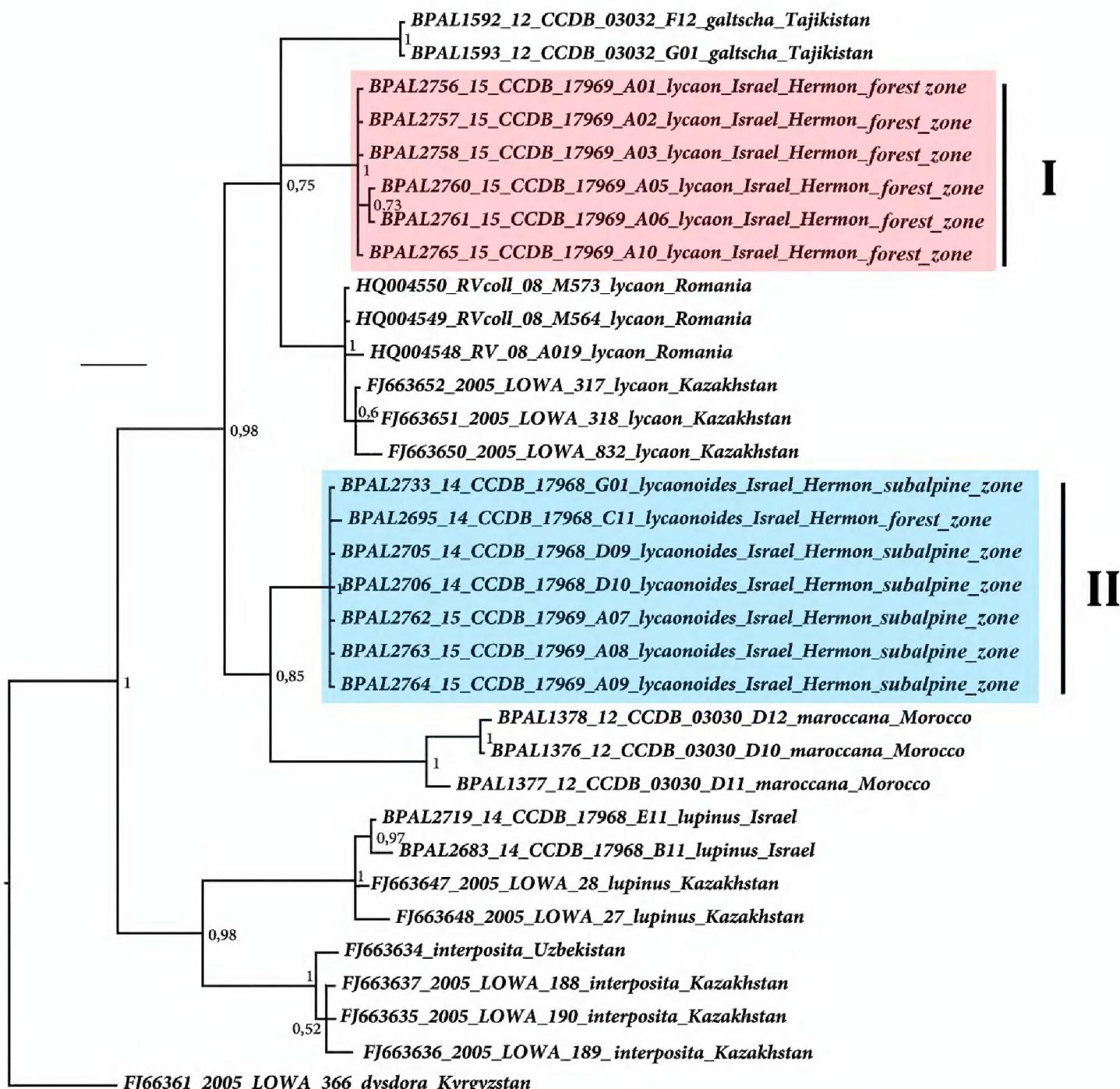


Figure 1. The Bayesian tree of the *Hyponephele lycaon* species complex based on analysis of *COI* DNA barcodes. Numbers at nodes indicate Bayesian posterior probability values. Sympatric haplogroups I and II from Israel are highlighted. Scale bar = 0.2 substitutions per position.

Table 2. Primary data (number of samples) for χ^2 -analysis of random vs. non-random distribution of the *COI* I and II haplogroups in the low (forest) and high (subalpine) zones.

	empirical values		expected values (in case of random distribution)	
	low altitude	high altitude	low altitude	high altitude
<i>COI</i> haplogroup I	6	0	3.234	2.772
<i>COI</i> haplogroup II	1	6	3.766	3.228

$df = 1$, 0.01 level of significance). Therefore, we reject the H_0 hypothesis and conclude that haplogroup I butterflies are significantly more frequent in the lower zone, whereas haplogroup II butterflies are significantly more frequent in the higher zone.



Figure 2. Biotopes on Mount Hermon, Israel where *COI* haplogroups I (a) and II (b) were collected.

The representatives of these two clusters were also different in the pattern on the hindwing underside (Figs 3 and 4). In haplogroup I this pattern had more contrast with clearly visible medial band, whereas in haplogroup II the hindwing underside was paler and had less contrast.

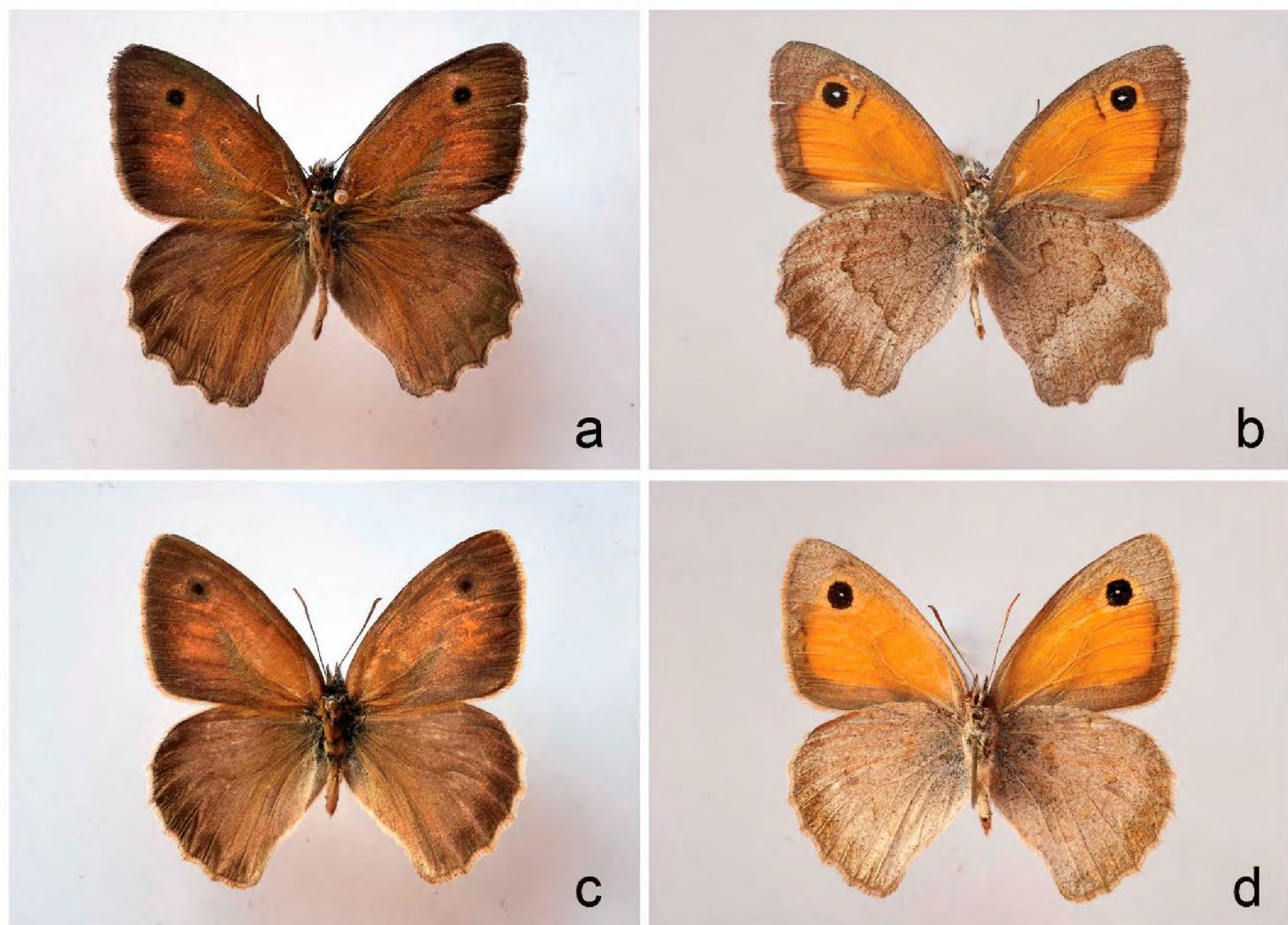


Figure 3. Wing pattern in haplogroup I and II samples from Mt Hermon, Israel. The pictures were taken using diffused daylight **a** sample CCDB-17969_A02, upperside **b** sample CCDB-17969_A02, underside **c** sample CCDB-17969_A09, upperside **d** sample CCDB-17969_A09, underside.

Table 3. Primary data (number of samples) for χ^2 -analysis of random *vs.* non-random association between the haplogroup I and II and the hindwing underside pattern.

	empirical values		expected values (in case of random distribution)	
	contrast pattern	pale	contrast pattern	pale
COI haplogroup I	6	0	2.772	3.234
COI haplogroup II	0	7	3.228	3.766

A standard χ^2 -test was used to distinguish between random *vs.* non-random association between haplogroups I and II and hindwing underside pattern (Table 3). The calculated χ^2 of 12.860 was larger than the tabular value (12.860 *vs.* 10.83, df = 1, 0.001 level of significance). Therefore, we reject the H_0 hypothesis and conclude that COI haplogroup I is significantly associated with contrast pattern of the hindwing underside, whereas COI haplogroup II is significantly associated with pale pattern of the hindwing underside.

The representatives of these two COI haplogroups also tended to be different in the length of the brachia in male genitalia (Fig. 5), although the latter character had high variability. Males of haplogroup I often had long brachia (Fig. 5a, b), whereas males of haplogroup II were mainly characterized by reduced brachia (Fig. 5c, d).

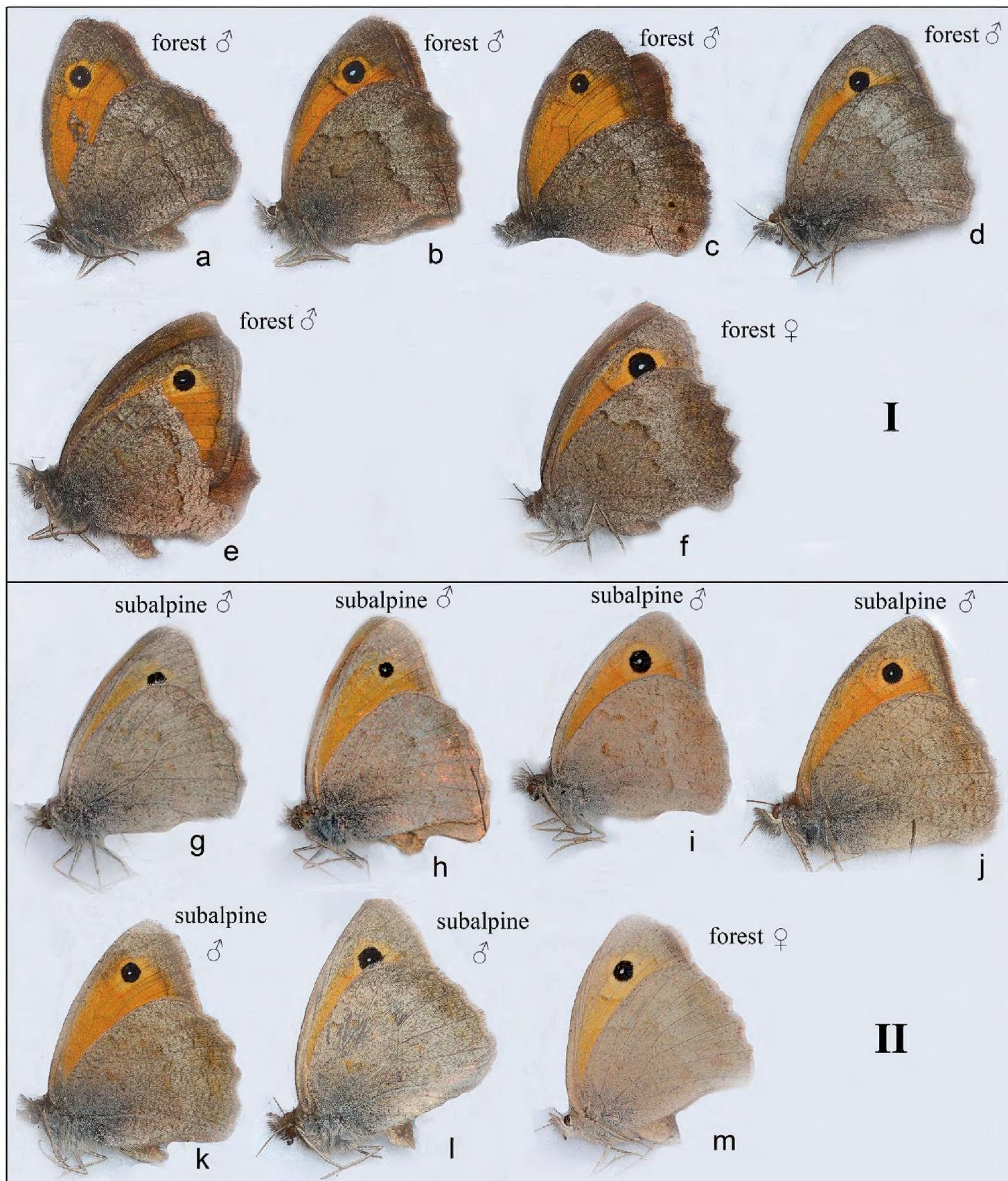


Figure 4. Pattern of the wing underside in haplogroups I and II samples. The pictures were taken using a flash **a** CCDB-17969_A01 **b** CCDB-17969_A02 **c** CCDB-17969_A03 **d** CCDB-17969_A06 **e** CCDB-17969_A10 **f** CCDB-17969_A05 **g** CCDB-17968_C11 **h** CCDB-17968_D09 **i** CCDB-17968_G01 **j** CCDB-17969_A07 **k** CCDB-17969_A08 **l** CCDB-17969_A09 **m** CCDB-17968_C11.

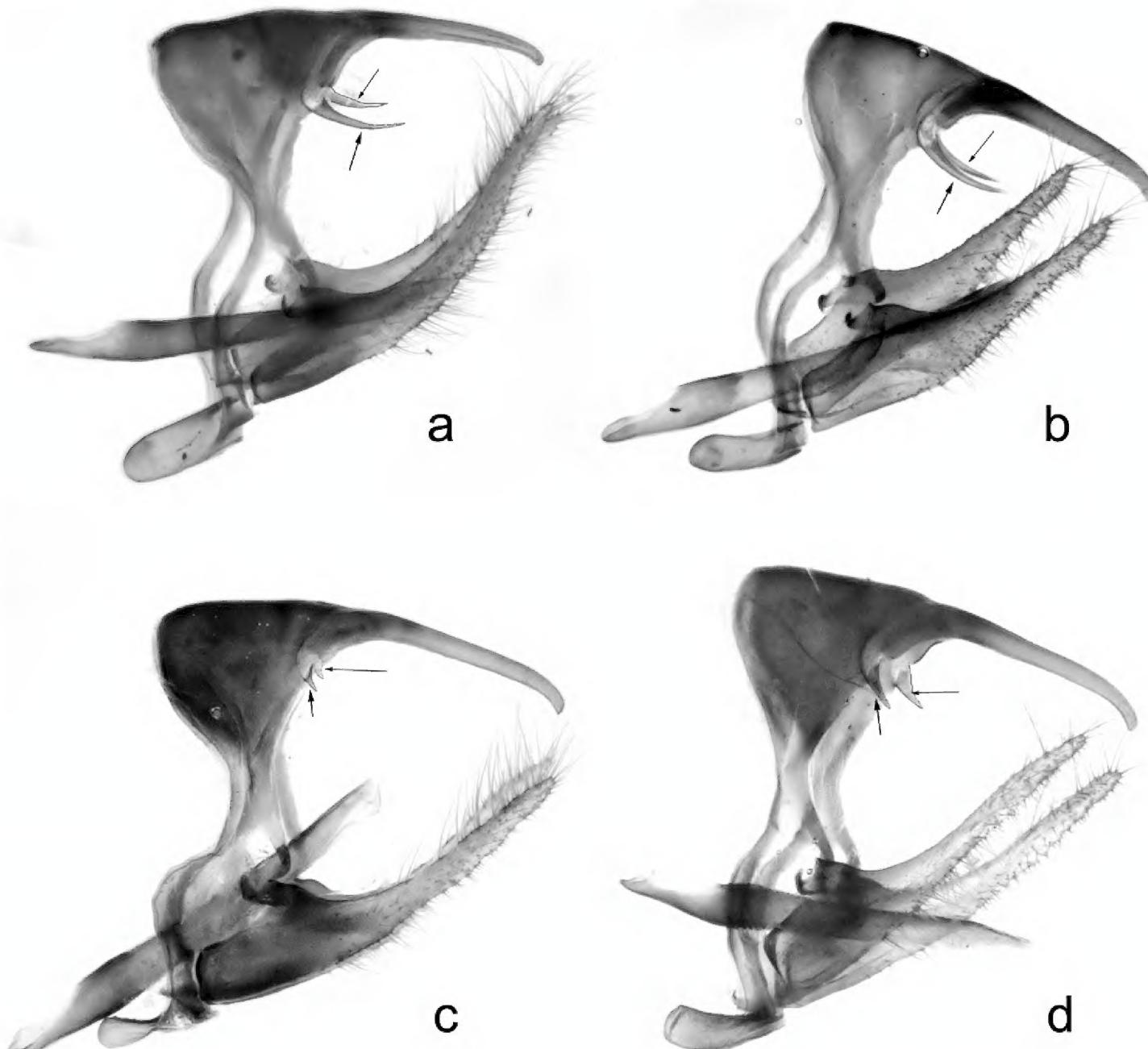


Figure 5. Typical male genitalia in haplogroups I (**a, b**) and II (**c, d**) from Mt Hermon, Israel. Lateral view. Brachia are indicated by arrow **a** specimen CCDB-17969_A10 **b** specimen CCDB-17969_A01 **c** specimen CCDB-17968_D10 **d** specimen CCDB-17968_D09.

Discussion

Evolutionary interpretation of the discovered pattern

The *COI* genetic distance between haplogroups I and II (3.5 %) is higher than the ‘standard’ 2.7–3.0% DNA-barcoding threshold commonly used as a tentative indicator for species distinctness of the taxa compared (Lambert et al. 2005, Lukhtanov et al. 2015a). It is known that *COI* barcodes alone are not sufficient for making any taxonomic decisions, since trees inferred from single markers sometimes display relationships that reflect the evolutionary histories of individual genes rather than the species being studied (Nichols 2001). Mitochondrial introgression (Zakharov et al. 2009) and *Wolbachia* infection (Ritter et al. 2013) can lead to additional bias when inferring taxonomic conclusions based on mitochondrial genes. Typically, multiple molecular markers or a combination of morphological and molecular markers are required for inferring taxonomic hypotheses. In our

research such additional information is represented by ecological characteristics (altitude belts). Less attention was attributed to the wing pattern, since we were not sure that it was an independent character. As the wing pattern strongly correlated with the ecology (low versus high elevation), one could hypothesize that the morphological difference is a consequence of phenotypic plasticity, i.e. ability of the same genotype to result in different phenotypes in response to changes in the environment (Price et al. 2003).

Three alternative explanations can account for bimodal sympatric distribution of mitochondrial markers. First, the diverged *COI* sequences may be selectively neutral intraspecific characters. Both preservation of a variety of ancestral haplotypes and mitochondrial introgression due to complex phylogeographic history could be responsible for such a neutral polymorphism (Avise 2000). Second, bimodal sympatric distribution of mitochondrial markers may be a result of a strong positive habitat-related selection working at intraspecific level and resulting in two *COI* clusters associated with different altitude belts (Cheviron and Brumfield 2009). Third, bearers of two diverged haplogroups may represent two different biological species (Avise 2000).

In our case the first hypothesis (neutral polymorphism) can be easily rejected. It predicts that the *COI* haplogroups I and II should be stochastically (i.e. randomly) distributed within high and low altitude belts. This prediction is not supported by χ^2 -test that demonstrated significantly non-random distribution of the *COI* haplogroups.

The second hypothesis (strong intraspecific positive selection) offers a more exotic, but not improbable, explanation. As *COI* sequence can be translated into a subunit of cytochrome c oxidase, a functional protein in mitochondria involved in energy metabolism (Kirk and Freedland 2011), this gene should be under natural selection (Castoe et al. 2008). Different haplotypes at this locus (or other linked mitochondrial genes) may be favoured in different environments. This could trigger a rapid sweep to fixation of a novel haplotype. This may result in sympatric clusters that differ in mitochondrial genes while exchanging alleles freely throughout the rest of the genome. Interestingly, such groups maintained by habitat-related selection could be considered species according to the genotypic cluster species concept (Coyne and Orr 2004, p. 448–449). The positive habitat-related selection of mitochondrial genome, despite its theoretical plausibility, has so far relatively low empirical support, although there are some data confirming mitochondrial evolution along temperature and altitude gradients (Cheviron and Brumfield 2009, Quintela et al. 2014).

The third hypothesis (two different species) seems to be a more likely explanation in the case of haplogroups I and II, especially if one takes into account the high level of genetic divergences between the haplogroups and concordance between molecular (Fig. 1), ecological (Fig. 2, Table 2) and morphological (Figs 3 and 4, Table 3) characters. More samples, especially from the intermediate elevation (1600–1800 m), and analysis of additional nuclear molecular markers across altitudinal transect will be required in future research to support or to reject the second (positive selection) and the third (two species) hypotheses and to reveal potential nuclear gene flow between haplogroups I and II.

Taxonomic interpretation of the discovered pattern

The presence of two sympatric, ecologically differentiated groups within *H. lycaon* complex in the Middle East is not a completely novel issue. A similar situation is known to exist in Iran and East Turkey (Weiss 1978, Eckweiler and Bozano 2011, Tshikolovets et al. 2014). It is accepted by *Hyponephele* genus experts (Eckweiler and Bozano 2011, Tshikolovets et al. 2014) that in Iran and Turkey these two groups represent two different species: *H. lycaon* and *H. lycaonoides* (but see the alternative opinion: Hesselbarth et al. 1995). Although we understand that this point of view requires an additional justification, we may accept it as a working hypothesis until further investigations and taxonomic revisions justify or falsify it.

If the species status of the discovered haplogroups will be confirmed in further studies, we suggest that, following Weiss (1978), the name *H. lycaon* (Rottenburg, [1775]) can be used for the Israeli taxon characterized by the contrast pattern on the hindwing underside and the predominance of longer brachia in male genitalia. Correspondingly, the name *H. lycaonoides* D. Weiss, 1978 can be used for the Israeli taxon characterized by the less contrasted pattern of the hindwing underside and the predominance of reduced brachia in male genitalia. However, this nomenclatural decision should be considered as a tentative one. First, despite recent revisions of the genus (Samodurov et al. 1995, 1996, 1997, 1999a, b, 2000, 2001, Eckweiler and Bozano 2011), no one studied type-specimens of numerous taxa that were described as subspecies and variations of *H. lycaon*. We cannot exclude that the name *lycaonoides* is a synonym of one of the previously described taxa, e.g. of *libanotica* (Staudinger, 1901). Second, molecular markers have never been used for analysis of taxonomic structure of *H. lycaon* species complex in its whole distribution range. Therefore, we will not be surprised if the true genetic and taxonomic structure of this group will be revealed as much more complex than a simple combination of two sympatric clusters as discovered in Iran, Turkey and Israel.

Acknowledgements

The financial support for this study was provided by the grant N 14-14-00541 from the Russian Science Foundation to the Zoological Institute of the Russian Academy of Sciences. We are grateful to Evgeny Zakharov (Canadian Centre for DNA Barcoding, Guelph, Canada) for sequencing the samples. We thank Dubi Benyamin for support of our DNA barcoding survey of Israeli butterflies and valuable comments.

References

- Avise JC (2000) Phylogeography: the history and formation of species. Harvard University Press, Cambridge, London, 447 pp.

- Benyamin D (2002) A Field Guide to the Butterflies of Israel. Keter Publishing House, Jerusalem, 248 pp.
- Castoe TA, Jiang ZJ, Gu W, Wang ZO, Pollock DD (2008) Adaptive evolution and functional redesign of core metabolic proteins in snakes. PLoS ONE 3(5): e2201. doi: 10.1371/journal.pone.0002201
- Cheviron ZA, Brumfield RT (2009) Migration-selection balance and local adaptation of mitochondrial haplotypes in rufous-collared sparrows (*Zonotrichia capensis*) along an elevational gradient. Evolution 63(6): 1593–1605. doi: 10.1111/j.1558-5646.2009.00644.x
- Coyne JA, Orr HA (2004) Speciation. Sinauer Associates, Sunderland, Massachusetts, 545 pp.
- deWaard JR, Ivanova NV, Hajibabaei M, Hebert PDN (2008) Assembling DNA barcodes: analytical protocols. In: Martin CC (Ed.) Environmental Genomics, Methods in Molecular Biology. Humana Press, Totowa, New Jersey 410: 275–283. doi: 10.1007/978-1-59745-548-0_15
- Dincá V, Zakharov EV, Hebert PD, Vila R (2011) Complete DNA barcode reference library for a country's butterfly fauna reveals high performance for temperate Europe. Proceedings of the Royal Society B 278(1704): 347–355. doi: 10.1098/rspb.2010.1089
- Eckweiler W, Bozano GC (2011) Guide to the butterflies of the Palearctic region. Satyrinae part IV. Tribe Satyrini, Subtribe Maniolina, *Maniola*, *Pyronia*, *Aphantopus*, *Hyponephele*. Omnes Artes, Milano, 102 pp.
- Hajibabaei M, deWaard JR, Ivanova NV, Ratnasingham S, Dooph RT, Kirk SL, Mackie PM, Hebert PDN (2005) Critical factors for assembling a high volume of DNA barcodes. Philosophical Transactions of the Royal Society of London. Series B, Biological Sciences 360: 1959–1967. doi: 10.1098/rstb.2005.1727
- Hall TA (1999) BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symposium Series 41: 95–98.
- Hesselbarth G, Oorchot H, Wagener S (1995) Die Tagfalter der Türkei unter Berücksichtigung der angrenzenden Länder. Selbstverlag Siegbert Wagener, Bocholt, Vol. 1–3, 1354 pp.
- Ivanova NV, deWaard JR, Hebert PDN (2006) An inexpensive, automation-friendly protocol for recovering high quality DNA. Molecular Ecology Resources 6: 998–1002. doi: 10.1111/j.1471-8286.2006.01428.x
- Kirk H, Freedland JR (2011) Applications and implications of neutral versus non-neutral markers in molecular ecology. International Journal of Molecular Sciences 12(6): 3966–3988. doi: 10.3390/ijms12063966
- Koçak AÖ (1989) *Hyponephele lycaonoides* Weiss in Turkey, with descriptions of new subspecies. Priamus 4: 142–146.
- Lambert DM, Baker A, Huynen L, Haddrath O, Hebert PDN, Millar CD (2005) Is a large-scale DNA-based inventory of ancient life possible? Journal of Heredity 96(3): 279–284. doi: 10.1093/jhered/esi035
- Lukhtanov VA, Shapoval NA, Dantchenko AV (2014) Taxonomic position of several enigmatic *Polyommatus* (*Agrodiaetus*) species (Lepidoptera, Lycaenidae) from Central and Eastern Iran: insights from molecular and chromosomal data. Comparative Cytogenetics 8(4): 313–322. doi: 10.3897/CompCytogen.v8i4.8939
- Lukhtanov VA, Dantchenko AV, Vishnevskaya MS, Saifitdinova AF (2015a) Detecting cryptic species in sympatry and allopatry: analysis of hidden diversity in *Polyommatus* (*Agrodiaetus*)

- butterflies (Lepidoptera: Lycaenidae). Biological Journal of the Linnean Society 116(2): 468–485. doi: 10.1111/bij.12596
- Lukhtanov VA, Sourakov A, Zakharov EV, Hebert PDN (2009) DNA barcoding Central Asian butterflies: increasing geographical dimension does not significantly reduce the success of species identification. Molecular Ecology Resources 9: 1302–1310. doi: 10.1111/j.1755-0998.2009.02577.x
- Lukhtanov VA, Shapoval NA, Anokhin BA, Saifitdinova AF, Kuznetsova VG (2015b) Homoploid hybrid speciation and genome evolution via chromosome sorting. Proceedings of the Royal Society B 282: 20150157. doi: 10.1098/rspb.2015.0157
- Nichols R (2001) Gene trees and species trees are not the same. Trends in Ecology and Evolution 16: 358–364. doi: 10.1016/S0169-5347(01)02203-0
- Posada D (2008) jModel Test: phylogenetic model averaging. Molecular Biology and Evolution 25(7): 1253–1256. doi: 10.1093/molbev/msn083
- Price TD, Qvarnström A, Irwin DE (2003) The role of phenotypic plasticity in driving genetic evolution. Proceedings of the Royal Society B 270(1523): 1433–1440. doi: 10.1098/rspb.2003.2372
- Quintela M, Johansson MP, Kristjánsson BK, Barreiro R, Laurila A (2014) AFLPs and mitochondrial haplotypes reveal local adaptation to extreme thermal environments in a freshwater gastropod. PLoS ONE 9(7): e101821. doi: 10.1371/journal.pone.0101821
- Ritter S, Michalski SG, Settele J, Wiemers M, Fric ZF, Sielezniew M, Šašić M, Rozier Y, Durka W (2013) *Wolbachia* infections mimic cryptic speciation in two parasitic butterfly species, *Phengaris teleius* and *P. nausithous* (Lepidoptera: Lycaenidae). PLoS ONE 8(11): 1–13. doi: 10.1371/journal.pone.0078107
- Ronquist F, Teslenko P, van der Mark D, Ayres A, Darling SH, Höhna B, Larget L, Liu M, Suchard A, Huelsenbeck JP (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61: 539–542. doi: 10.1093/sysbio/sys029
- Samodurov GD, Korolew WA, Tshikolovets VV (1996) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 2. Atalanta 27: 223–252.
- Samodurov GD, Korolew WA, Tshikolovets VV (1997) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 3. Atalanta 28: 49–96.
- Samodurov GD, Korolew WA, Tshikolovets VV (1999a) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 4. Atalanta 29: 25–68.
- Samodurov GD, Korolew WA, Tshikolovets VV (1999b) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 5. Atalanta 29: 69–105.
- Samodurov GD, Korolew WA, Tshikolovets VV (2000) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 6. Atalanta 31: 135–170.
- Samodurov GD, Korolew WA, Tshikolovets VV (2001) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. 7. Atalanta 32: 111–186.
- Samodurov GD, Tshikolovets VV, Korolew WA (1995) Eine Übersicht über die Satyriden der Gattung *Hyponephele* Mushamp, 1915. I. Atalanta 26: 157–195.

- Talavera G, Lukhtanov V, Rieppel L, Pierce NE, Vila R (2013) In the shadow of phylogenetic uncertainty: the recent diversification of *Lysandra* butterflies through chromosomal change. *Molecular Phylogenetics and Evolution* 69: 469–478. doi: 10.1016/j.ympev.2013.08.004
- Tshikolovets V, Naderi A, Eckweiler W (2014) The Butterflies of Iran and Iraq. Tshikolovets Publications, Pardubice, 440 pp. <http://www.entosphinx.cz/985-1202-thickbox/>
- Vershinina AO, Lukhtanov VA (2010) Geographical distribution of the cryptic species *Agrodiaetus alcestis alcestis*, *A. alcestis karacetinae* and *A. demavendi* (Lepidoptera, Lycaenidae) revealed by cytogenetic analysis. *Comparative Cytogenetics* 4(1): 1–11. doi: 10.3897/compcytogen.v4i1.21
- Weiss D (1978) A new species of the genus *Hyponephele* Muschamp 1915 from West Iran. *Atalanta* 9: 230–232.
- Zakharov EV, Lobo NF, Nowak C, Hellma JJ (2009) Introgression as a likely cause of mtDNA paraphyly in two allopatric skippers (Lepidoptera: Hesperiidae). *Heredity* 102: 590–599. doi: 10.1038/hdy.2009.26